

Development and validation of analytical RP-HPLC method for the quantitative determination of avapritinib in pure substances and marketed pharmaceutical tablet forms

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Abstract

The present work includes a simple, economic, rapid, accurate and precise isocratic RP-HPLC method development for estimation of Avapritinib in bulk form and its marketed formulation. Estimation was done at 286nm which was found to be λ max of Avapritinib. The simple, selective, isocratic RP-HPLC method for Avapritinib was developed on Phenomenex Luna (C₁₈) RP Column; 250 mm x 4.6 mm, 5µm with a mobile phase of Phosphate Buffer (pH-4.6) and Methanol were taken in the ratio of 65:35% v/v at a flow rate of 1.0 ml/min and detection wavelength 286nm. The developed method was validated successfully according to ICH Q2 (R1) guidelines. The chromatographic methods showed a good linear response with r2 values of 0.9995. The percentage relative standard deviation for method was found to be less than two, indicating that the methods were precise. The mean percentage recovery was for RP-HPLC method was 100.437%. From the results it could be concluded that both the developed method was specific, selective and robust. The method could be successfully applied for analysis of Bulk form and Marketed formulation of Avapritinib.

Keywords: Avapritinib, RP-HPLC, Method Development, Validation, ICH Guidelines

Introduction

Avapritinib, or BLU-285, is a selective tyrosine kinase inhibitor of KIT and platelet derived growth factor receptor alpha indicated for the treatment of unresectable, metastatic gastrointestinal stromal tumours ^[1]. It is one of the first medications available for the treatment of multidrug resistant cancers. Avapritinib shares a similar mechanism with [Ripretinib]. Avapritinib was granted FDA approval on 9 January 2020. Avapritinib is a selective kinase inhibitor that negatively modulates the action of cell transporters to resensitize them to other chemotherapies. It has a long duration of action as it is given once daily.3 Patients should be counseled regarding the risk of intracranial hemorrhage, CNS effects, and embryo-fetal toxicity. Avapritinib has a negative modulating effect on the transporters ABCB1 and ABCG2, which mediate the multidrug resistance phenotype of some cancers ^[2]. This modulation may be due to interactions of Avapritinib with the drug binding pocket of these transporters. Negative modulation of these transporters, resensitizes cancerous cells to treatment with chemotherapeutic agents like paclitaxel ^[3]. The IUPAC name of Avapritinib is (1S)-1-(4-fluorophenyl)-1-[2-[4-[6-(1-methylpyrazol-4-yl)pyrrolo[2,1-f][1,2,4]triazin-4-yl]piperazin-1-yl]pyrimidin-5-yl] ethanamine. The Chemical Structure of Avapritinib shown in follows.



Fig 1: Chemical Structure of Avapritinib

Materials and Methods

Table 1: List of Instrument used

| S. No. | Instruments/Equipments/Apparatus |
|--------|---|
| 1. | HPLC with Empower2 Software with Isocratic with UV-Visible Detector (Waters). |
| 2. | ELICO SL-159 UV – Vis spectrophotometer |
| 3. | Electronic Balance (SHIMADZU ATY224) |
| 4. | Ultra Sonicator (Wensar wuc-2L) |
| 5. | Thermal Oven |
| 6. | Symmetry ODS RP C ₁₈ ,5µm, 15mm x 4.6mm i.d. |
| 7. | P ^H Analyzer (ELICO) |
| 8. | Vacuum filtration kit (BOROSIL) |

Table 2: List of Chemicals Used

| C No | Numera | Specifications | | Manufa atoma /Some Har | |
|---------|-------------------------|----------------|-------|--------------------------|--|
| 5. INO. | Name | Purity | Grade | Manufacturer/Supplier | |
| 1. | Doubled distilled water | 99.9% | HPLC | Sd fine-Chem ltd; Mumbai | |
| 2. | HPLC Grade Water | 99.9% | HPLC | Sd fine-Chem ltd; Mumbai | |
| 3. | Methanol | 99.9% | HPLC | Loba Chem; Mumbai. | |
| 4. | Hydrochloric Acid | 99.9 | A.R. | Sd fine-Chem ltd; Mumbai | |
| 5. | Acetonitrile | 99.9% | HPLC | Loba Chem; Mumbai. | |
| 6. | Sodium Hydroxide | 99.9 | A.R. | Sd fine-Chem ltd; Mumbai | |
| 7. | Ethanol | 99.9 | A.R. | Sd fine-Chem ltd; Mumbai | |
| 8. | Octanol | 99.9 | A.R. | Sd fine-Chem ltd; Mumbai | |

Optimization of Chromatographic Conditions: The chromatographic conditions ^[4-6] was optimized by different means. (Using different column, different mobile phase,

different flow rate, different detection wavelength & different diluents for sample preparation etc.

|--|

| Column Used | Mobile Phase | Flow Rate | Wave length | Observation | Result |
|------------------------------------|------------------------------|--------------|-------------|-------------------------|----------|
| Xterra ODS (C18) RP Column, 150 | Methanol: Acetonitrile: | 0.8m1/min | 286nm | Voru Low response | Method |
| mm x 4.6 mm, 5µm | Water = 40:30:30 | 0.8111/11111 | 2801111 | Very Low response | rejected |
| Zorbax ODS (C18) RP Column, 250 | Methanol: Acetonitrile = 70: | 0.0m1/min | 286nm | Low response | Method |
| mm x 4.6 mm, 5µm | 30 | 0.9111/11111 | 2801111 | Low response | rejected |
| Develosil ODS (C18) RP Column, 150 | Acetonitrile: Methanol = 60: | 1.0ml/min | 286nm | Tailing paaks | Method |
| mm x 4.6 mm, 5µm | 40 | 1.0111/11111 | 2001111 | Tannig peaks | rejected |
| Phenomenex Luna (C18) RP Column, | Phosphate Buffer (pH-5.2): | 1.0m1/min | 296000 | Desclution was not good | Method |
| 250 mm x 4.6 mm, 5µm | Methanol = 80:20 | 1.0111/11111 | 2001111 | Resolution was not good | rejected |
| Phenomenex Luna (C18) RP Column, | Phosphate Buffer (pH-3.8): | 1.0ml/min | 286nm | Tailing paak | Method |
| 250 mm x 4.6 mm, 5µm | Methanol = 55:45 | 1.0111/11111 | 2001111 | Taning peak | rejected |
| Phenomenex Luna (C18) RP Column, | Phosphate Buffer (pH-4.6): | 1.0ml/min | 286nm | Nice peak | Method |
| 250 mm x 4.6 mm, 5µm | Methanol = 65:35 | 1.0111/11111 | 2001111 | Nice peak | accepted |

Preparation of Mobile Phase: 650ml of prepared phosphate buffer and 350ml of HPLC Grade Methanol were mixed well and degassed in ultrasonic water bath for 15 minutes. The solution was filtered $^{[7]}$ through 0.45 μm filter under vacuum filtration.

Analytical Method Validation

Method validation is the process used to confirm that the analytical procedure employed for a specific test is suitable for its intended use. Results from method validation ^[8-10] can be used to judge the quality, reliability and consistency of analytical results; it is an integral part of any good analytical practice.

System Suitability Parameters: The theory of chromatography has been used as the basis for system-suitability tests, which are set of quantitative criteria that test the suitability of the chromatographic system to identify and quantify drug related samples by HPLC at any step of the pharmaceutical analysis.

Specificity: Specificity is the ability to assess unequivocally the analyte in the presence of components which may be expected to be present. Typically these might include impurities, degradants, matrix, etc. Lack of specificity of an individual analytical procedure may be compensated by other supporting analytical procedure(s).

Accuracy: The accuracy of an analytical procedure expresses the closeness of agreement between the value which is accepted either as a conventional true value or an accepted reference value and the value found. This is sometimes termed trueness.

Precision: The precision of an analytical procedure expresses the closeness of agreement (degree of scatter) between a series of measurements obtained from multiple sampling of the same homogeneous sample under the prescribed conditions. Precision may be considered at three levels: repeatability, intermediate precision and reproducibility. Precision should be investigated using homogeneous, authentic samples.

Repeatability: Repeatability expresses the precision under the same operating conditions over a short interval of time. Repeatability is also termed intra-assay precision.

Intermediate precision: Intermediate precision expresses within-laboratories variations: different days, different analysts, different equipment, etc.

Reproducibility: Reproducibility expresses the precision between laboratories (collaborative studies, usually applied to standardization of methodology).

Detection Limit: The detection limit of an individual analytical procedure is the lowest amount of analyte in a

sample which can be detected but not necessarily quantitated as an exact value.

Quantitation Limit: The quantitation limit of an individual analytical procedure is the lowest amount of analyte in a sample which can be quantitatively determined with suitable precision and accuracy. The quantitation limit is a parameter of quantitative assays for low levels of compounds in sample matrices, and is used particularly for the determination of impurities and/or degradation products.

Linearity: The linearity of an analytical procedure is its ability (within a given range) to obtain test results which are directly proportional to the concentration (amount) of analyte in the sample.

Range: The range of an analytical procedure is the interval between the upper and lower concentration (amounts) of analyte in the sample (including these concentrations) for which it has been demonstrated that the analytical procedure has a suitable level of precision, accuracy and linearity.

Robustness: The robustness of an analytical procedure is a measure of its capacity to remain unaffected by small, but deliberate variations in method parameters and provides an indication of its reliability during normal usage.

Results and Discussion Development of Analytical Method Selection of Wavelength

The standard & sample stock solutions were prepared separately by dissolving standard & sample in a solvent in mobile phase ^[11] diluting with the same solvent. (After optimization of all conditions) for UV analysis. Itscanned in the UV spectrum in the range of 200 to 400nm. This has been performed to know the maxima of Avapritinib, so that the same wave number can be utilized in HPLC UV detector for estimating the Avapritinib. The scanned UV spectrum is attached in the following page,

Sample & Standard Preparation for the UV-Spectrophotometer Analysis: 25 mg of Avapritinib standard ^[12] was transferred into 25 ml volumetric flask, dissolved & make up to volume with mobile phase. Further dilution was done by transferring 1ml of the above solution into a 10ml volumetric flask and make up to volume with mobile phase.



Fig-2: UV Spectrum for Avapritinib

Observation: While scanning the Avapritinib solution we observed the maxima at 286nm. The UV spectrum ^[13] has

been recorded on ELICO SL-159 make UV – Vis spectrophotometer model UV-2450.

Optimization of Method Summary of Optimized Chromatographic Conditions:

The Optimum Chromatographic conditions ^[14-16] obtained from experiments can be summarized as below:

| Table 4: Summary of optimized | I Chromatographic Conditions |
|-------------------------------|------------------------------|
|-------------------------------|------------------------------|

| Mobile phase | Phosphate Buffer (pH-4.6): Methanol = 65:35% |
|-----------------------------|---|
| Column | Phenomenex Luna (C18) RP Column, 250 mm x 4.6 mm, 5µm |
| Column Temperature | Ambient |
| Detection Wavelength | 286 nm |
| Flow rate | 1.0 ml/ min. |
| Run time | 10 min. |
| Temperature of Auto sampler | Ambient |
| Diluent | Mobile Phase |
| Injection Volume | 20µ1 |
| Type of Elution | Isocratic |
| Retention time | 4.817 minutes |



Fig 3: Chromatogram for Blank Solution



Fig 4: Chromatogram of Avapritinib in Optimized Condition

Observation: The selected and optimized mobile phase was Phosphate Buffer (pH-4.6): Methanol = 65:35% and conditions optimized were flow rate (1.0 ml/minute), wavelength (286nm), Run time was 10 mins. Here the peaks were separated and showed better resolution, theoretical plate count and symmetry ^[17]. The proposed chromatographic conditions were found appropriate for the quantitative determination of the drug.

Validation of Analytical Method 1. System Suitability Test

System suitability testing is an integral part of many analytical procedures. The tests are based on the concept that the equipment, electronics, analytical operations and samples to be analysed constitute an integral system that can be evaluated as such. Following system suitability test parameters ^[18-19] were established. The data are shown in Table-5 & 6.

| S. No. | Injection No. | RT | Area | USP Plate Count | USP Tailing |
|--------|---------------|-------|-------------|-----------------|-------------|
| 1 | Injection 1 | 4.817 | 745236 | 6986 | 1.39 |
| 2 | Injection 2 | 4.783 | 743652 | 6857 | 1.37 |
| 3 | Injection 3 | 4.840 | 742587 | 6856 | 1.36 |
| 4 | Injection 4 | 4.783 | 742946 | 6847 | 1.39 |
| 5 | Injection 5 | 4.817 | 743654 | 6896 | 1.38 |
| 6 | Injection 6 | 4.778 | 741698 | 6874 | 1.37 |
| Mean | | | 743295.5 | 6886 | 1.37666 |
| S.D | | | 1199.773604 | | |
| %RSD | | | 0.161412736 | | |

Table 5: Data of System Suitability Test

Table 6: Data of System Suitability Parameter

| S. No. | Parameter | Limit | Result |
|--------|-------------------|------------|--------------------|
| 1 | Retention Time | RT> 2 | Avapritinib= 4.778 |
| 2 | Asymmetry | $T \leq 2$ | Avapritinib= 1.35 |
| 3 | Theoretical plate | N > 2000 | Avapritinib= 6859 |
| 4 | Tailing Factor | T<2 | Avapritinib= 1.37 |

2. Linearity

To evaluate the linearity, serial dilution of analyte were prepared from the stock solution was diluted with mobile phase to get a series of concentration ranging from 60- 140μ g/ml. The prepared solutions were sonicated. From these

solutions, 10μ l injections of each concentration were injected into the HPLC system and chromatographed under the optimized conditions. Calibration curve ^[20] was constructed by plotting the mean peak area (Y-axis) against the concentration (X-axis).



Fig 5: Calibration Curve of Avapritinib

Table 7: Linearity Data for Avapritinib

| Conc. (µg/ml) | Area |
|---------------|---------|
| 0 | 0 |
| 60 | 461404 |
| 80 | 606157 |
| 100 | 748506 |
| 120 | 891041 |
| 140 | 1032196 |

3. Accuracy: The accuracy of the method was determined by recovery studies and the percentage recovery ^[21] was calculated. The recoveries of Avapritinib were found to be in the range of 99-102%. The proposed Liquid Chromatographic method was applied to the determination of Avapritinib. The results for Avapritinib comparable with the corresponding labeled amounts.

| Accuracy | Amount Added | Amount Recovered | Peak Area | % Recovery | Mean Recovery |
|----------|--------------|------------------|-----------|------------|---------------|
| | 80 | 80.798 | 604517 | 100.997 | |
| 80% | 80 | 80.673 | 603598 | 100.841 | |
| | 80 | 80.756 | 604213 | 100.945 | 100 42704 |
| | 100 | 99.933 | 745471 | 99.933 | 100.437% |
| 100% | 100 | 100.083 | 746574 | 100.083 | |
| | 100 | 100.365 | 748652 | 100.365 | |

| | 120 | 120.290 | 895415 | 100.241 |
|------|-----|---------|--------|---------|
| 120% | 120 | 120.201 | 894762 | 100.167 |
| | 120 | 120.442 | 896541 | 100.368 |

4. Precision

Repeatability: The precision ^[22] of each method was ascertained separately from the peak areas & retention times obtained by actual determination of six replicates of a fixed

amount of drug. Avapritinib (API). The percent relative standard deviation was calculated for Avapritinib are presented in the table-9.

| S. No. | Injection | Peak Area |
|--------|-------------|-------------|
| 1 | Injection 1 | 743826 |
| 2 | Injection 2 | 745277 |
| 3 | Injection 3 | 742506 |
| 4 | Injection 4 | 747576 |
| 5 | Injection 5 | 746715 |
| 6 | Injection 6 | 741278 |
| 7 | Average | 744529.6667 |
| 8 | SD | 2440.4116 |
| 9 | % RSD | 0.32777 |

Table 9: Repeatability Data for Avapritinib

Intermediate Precision:

The Intermediate Precision ^[23-25] consists of two methods:-**Intra Day:** In Intra Day process, the 80%, 100% and 120% concentration are injected at different intervals of time in same day.

Inter Day:In Inter Day process, the 80%, 100% and 120% concentration are injected at same intervals of time in different days.

Table 10: Results of intra-assay & inter-assay

| | Observed Conc. of Avapritinib (µg/ml) by the proposed method | | | |
|-----------------------------------|--|-------|------------|-------|
| Conc. of Avapritinib(API) (µg/ml) | Intra-Day | | Inter-Day | |
| | Mean (n=6) | % RSD | Mean (n=6) | % RSD |
| 80 | 80.096 | 0.487 | 79.685 | 0.688 |
| 100 | 100.074 | 0.968 | 100.057 | 0.789 |
| 120 | 120.056 | 0.847 | 120.016 | 0.698 |

Observations: The intra & inter day variation of the method was carried out for standard deviation & % RSD (% RSD < 2%) within a day & day to day variations for Avapritinib revealed that the proposed method is precise ^[26].

5. Specificity

Specificity can be determined by comparing the chromatograms obtained from the drugs with the chromatogram obtained from the blank solution. Blank solution was prepared by mixing the excipients in the mobile phase without drug. Drug solutions were prepared individually and the sample containing one drug was also prepared. Now these mixtures were filtered by passing through 0.45 μ membrane filter before the analysis. In this observation no excipient peaks were obtained near the drug in the study run time. This indicates that the proposed method was specific ^[28].

Observation: In this test method blank, standard solutions were analyzed individually to examine the interference. The abovechromatograms show that the active ingredient was well separated from blank and their excipients and there was no interference of blank with the principal peak. Hence the method is specific.

6. Limit of Detection (LOD) and Limit of Quantification (LOQ)

The LOD ^[29] and LOQ parameter was evaluated by mistreatment the slope of line and variance obtained from accuracy studies.

The detection limit (LOD) and quantization limit (LOQ) may be expressed as:

L.O.D. = 3.3(SD/S) L.O.Q. = 10(SD/S)

Where, SD = Standard deviation of the response S = Slope of the calibration curve

The slope S may be estimated from the calibration curve of the analyte.

The Minimum concentration level at which the analyte can be reliable detected (LOD) & quantified ^[30] (LOQ) were found to be $1.469 \& 4.454 \mu g/ml$ respectively.

7. Method Robustness: Influence of small changes in chromatographic conditions such as change in flow rate 1.0 ml (\pm 0.1 ml/min), Wavelength of detection 286 (\pm 2nm) & organic phase content in mobile phase (\pm 5%) studied to determine the robustness ^[31] of the method are also in favour of (Table-11, % RSD < 2%) the developed RP-HPLC method for the analysis of Avapritinib (API).

Effect of Variation of Flow Conditions

recorded.

The sample was analyzed at 0.9 ml/min and 1.1 ml/min instead of 1ml/min, remaining conditions are same. 10μ l of the above sample was injected and chromatograms were recorded.

Effect of Variation of Mobile Phase Organic Composition The sample was analyzed by variation of mobile phase i.e. Methanol: Phosphate Buffer was taken in the ratio and 40:60, 30:70 instead of 35:65, remaining conditions are same. 20µl of the above sample was injected and chromatograms were

| Parameter Used for Sample Analysis | Peak Area | Retention Time | Theoretical Plates | Tailing Factor |
|------------------------------------|-----------|-----------------------|---------------------------|-----------------------|
| Actual Flow rate of 1.0 mL/min | 742946 | 4.778 | 1.37 | 2896 |
| Less Flow rate of 0.9 mL/min | 698965 | 4.783 | 1.39 | 2986 |
| More Flow rate of 1.1 mL/min | 786598 | 4.817 | 1.42 | 2985 |
| Less organic phase | 732642 | 4.842 | 1.29 | 3102 |
| More organic phase | 702546 | 4.773 | 1.37 | 3247 |

Table 11: Results for Robustness

8. Estimation of Avapritinib in Pharmaceutical Dosage Form

Label Claim: 100mg

Each tablet contains: 100mg

Twenty Tablets were taken and the I.P. method was followed to determine the average weight. Above weighed tablets were finally powdered and triturated well. A quantity of powder equivalent to 25 mg of drugs were transferred to 25 ml volumetric flask, make and solution was sonicated for 15 minutes, there after volume was made up to 25 ml with same solvent. Then 10ml of the above solution was diluted to 100 ml with mobile phase. The solution was filtered through a membrane filter (0.45 μ m) and sonicated to degas. The solution prepared was injected in five replicates into the HPLC system and the observations were recorded. The Assay ^[32] data are shown in Table-12.

Assay

Assay % =

$$\frac{AT}{AS} \times \frac{WS}{DS} \times \frac{DT}{WT} \times \frac{P}{100} \times Avg Wt = mg$$

Where:

AT = Peak Area of drug obtained with test preparation AS = Peak Area of drug obtained with standard preparation WS = Weight of working standard taken in mg WT = Weight of sample taken in mg DS = Dilution of Standard solution

DT = Dilution of sample solution

P = Percentage purity of working standard

| Table 12: Recovery | / Data for Estimation | n Avapritinib | in Ayvakit Tablet |
|--------------------|-----------------------|---------------|-------------------|
|--------------------|-----------------------|---------------|-------------------|

| Brand name of Avapritinib | Labelled amount of | Mean (± SD) amount (mg) found | Assay % |
|--------------------------------------|--------------------|-------------------------------|-----------------|
| | Drug (mg) | by the proposed method (n=6) | (± SD) |
| Ayvakit Tablet (Blueprint Medicines) | 100mg | 99.685 (± 0.698) | 99.743 (±0.268) |

Result & Discussion: The amount of drug in Ayvakit Tablet was found to be 99.685 (± 0.698) mg/tab for Avapritinib& %Purity was 99.743%.

Summary and Conclusion

To develop a precise, linear, specific & suitable stability indicating RP-HPLC method for analysis of Avapritinib, different chromatographic conditions were applied & the results observed are presented in previous chapters. Isocratic elution is simple, requires only one pump & flat baseline separation for easy and reproducible results. So, it was preferred for the current study over gradient elution. In case of RP-HPLC various columns are available, but here Phenomenex Luna (C18) RP Column, 250 mm x 4.6 mm, 5µmColumn was preferred because using this column peak shape, resolution and absorbance were good. Detection wavelength was selected after scanning the standard solution of drug over 200 to 400nm. From the U.V spectrum of Avapritinib it is evident that most of the HPLC work can be accomplished in the wavelength range of 286 nm conveniently. Further, a flow rate of 1.0 ml/min & an injection volume of 20µl were found to be the best analysis. The result shows the developed method is yet another suitable method for assay and stability related impurity studies which can help in the analysis of Avapritinib in different formulations.

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