



Influence of inorganic, organic and bio fertilizer on chlorophyll content, relative water and EC content of leaf of host plant of Tasar silkworm (*Antheraea mylitta* Drury)

Soma Karmakar^{1*}, AK Srivastava², T Pandiaraj³

¹⁻² University Department of Botany, Ranchi University, Ranchi, India

³ Narendra Deva University of Agricultural Technology, Azamgarh, Uttar Pradesh, India

* Corresponding Author: Soma karmakar

Article Info

ISSN (online): 2582-7138

Volume: 04

Issue: 02

March-April 2023

Received: 04-03-2023;

Accepted: 23-03-2023

Page No: 383-388

Abstract

The research was done to find out the influence of Inorganic, Organic and Biofertilizer on Chlorophyll content, Relative water and EC content of leaf of host plant of Tasar Silkworm (*Antheraea mylitta* Drury). The results showed that application of different combination of manure significantly influenced the Chlorophyll content, Relative water and EC content. The RWC was found highest in T₁₇(85.2%) was applied with 100% RDF through fertilizer +2kg vermicompost+ PSB followed by T₁₈(85.02%) and T₃(84.99%). The EC was found highest in T₁₉(0.09405 dSm⁻¹) followed by T₁₈(0.0922 dSm⁻¹). After 10 min the EC was found to be highest in T₁₉(0.245dSm⁻¹) followed by T₆(0.230dSm⁻¹). The Chl 'a' was recorded high in T₁₆(4.70mg/g), Chl 'b' in T₇(3.18 mg/g) and total chlorophyll was recorded highest in T₇(7.68 mg/g) followed by T₁₆(7.55 mg/g). The obtained results showed that combined application of manure have significant affect on chlorophyll content, Relative water and EC content of leaf.

Keywords: Biofertilizer, Chlorophyll, Relative water, Silkworm and Vermicompost

Introduction

Silk farming is a commercial cultivation providing valuable foreign exchange (www.imotforum.com) and employment and livelihood to approximately 9.4 million persons in rural and semi-urban areas. It is the oldest industry in the country. All the five known commercial silks namely Mulberry, Tropical Tasar, Oak-Tasar, Eri and Muga are produced by India making it unique in the world (Central Silk board). Good quality of Silk production depend on plant nutrition. For optimal growth of plant, nutrients must be present in sufficient and balanced quantities. Fertilization of the soil are done to maintain the natural fertility and which also act as a sources of plant nutrient (Purwanto and Junaidah, 2015) [26]. Use of organic fertilizer can meet the nutrient requirements for sustainable production. Management of organic and biofertilizer reduces the potential damage than mineral fertilizer (Ahmed *et al.*, 2011) [1]. Biofertilizer application in agriculture is a sustainable way of increasing crop yields and reduces production cost (Wali Asal, 2010) [37]. Bio-fertilization is a safe process for human, animal and environments which lower pollution and fertilization cost. It helps in improving the soil biota and also reduces the sole use of chemical fertilizers (Sabashini *et al.*, 2007) [29]. Organic material act as soil conditioners, they not only serve as source of plant nutrients but also improve soil physical properties, as evidenced by increased water infiltration, water holding capacity, aeration and permeability, soil aggregation and rooting depth and by decreasing soil crusting, bulk density and erosion (Allision 1973, USDA 1978). Application of organic and inorganic fertilizers to the soil helps in growth of plants by supplying nutrients and thus affects the physiological processes of plants (Amujoyegbe *et al.*, 2007) [4].

RWC indicate the plant water status and was used instead of plant water potential. It indicate the balance between absorbed water by plant and consumed through transpiration (M. Hassanzadeh *et al.*, 2009) [20]. Plant having high RWC, are more resistant against drought stress (Schonfeld *et al.*, 1988) [30]. Photosynthetic capacity decreases usually at relative water content below 70% which in many plants correspond to severe wilting of leaves but these inhibitory effects are still reversible down to 30%-70% of relative water content (Kaiser, 1987) [15].

EC is related to amount of ions available in the root zone of plant (Nemail and Van Iersel, 2004) [24]. The salt concentration and electrolyte concentration index of the solution are determined by EC (Kozai *et al.*, 2015; Lu *et al.*, 2017) [16, 19]. The EC depend on the environmental conditions and for different crop it has different optimal EC (Le Bot *et al.*, 1998; Sonneveld and Voogt, 2009) [17, 34]. Higher EC causes environmental pollution by increasing the discharge of nutrients into the environment and also hinder nutrient uptake by increasing the osmotic pressure. Plant health and yield are severely affected by lower EC (Signore *et al.*, 2016; Lu *et al.*, 2017) [32, 19].

Chlorophylls in the plant are the most important green pigments for photosynthetic process (Bhatia and parashar, 1997) [7]. Five types of chlorophyll occur in plants a, b, c, d and e out of these, in higher plant two chlorophyll i.e. a and b are found. chlorophyll a is termed as universal photosynthetic pigment and also known as primary photosynthetic pigment because it convert light into chemical or electrical energy which is the primary reaction centre of photosynthesis. Other pigments are accessory pigments. They absorb light energy of different wavelengths and pass the energy to chlorophyll a through electron spin resonance (Bhatia and Tyagi, 2006) [6].

Leaf chlorophyll content is indicator of photosynthetic capability of plant tissue (Nageswara *et al.* 2001; Write *et al.*, 1994) [23] and also provide indirect but accurate estimate of nutritional status of plant (Vijay paul *et al.*, 2017) [36]. Chlorophyll green coloration is related to the amount of nutrient absorbed by the plant from soil (Roy and Singh, 2006) and also observed that fertilization with organic manure increase chlorophyll content than inorganic fertilizer. Photosynthetic activity and crop yield increase with increased chlorophyll content in leaf (Siavoshi and Laware, 2013) [31]. This research was done to find the influence of inorganic, organic and biofertilizer on chlorophyll content, Relative water and EC content of leaf of host plant of Tasar Silkworm (*Antheraea mylitta* Drury).

Materials and Methods

The present study was carried out in the field of Central Tasar Research & Training institute, Nagri, Ranchi to determine the influence of inorganic, organic and biofertilizer on Chlorophyll content, Relative water and EC content of leaf of host plant of Tasar Silkworm (*Antheraea mylitta* Drury). Twenty different combination with three replication were laid out in Randomized Complete Block Design (Table1).

Table1: Treatment Details

Treatments	Treatments detail
T ₁	Absolute Control
T ₂	Control with recommended dose fertilizer(RDF)
T ₃	50% RDF through fertilizer+50% through vermicompost
T ₄	75% RDF through fertilizer+25% through vermicompost
T ₅	100% RDF through fertilizer+2% through vermicompost
T ₆	50%RDF+Azotobacter
T ₇	75%RDF+Azotobacter
T ₈	100%RDF+Azotobacter
T ₉	50% RDF+ Phosphorus solubilizing bacteria(PSB)
T ₁₀	75%RDF+PSB
T ₁₁	100%RDF+PSB
T ₁₂	T ₃ +Azotobacter
T ₁₃	T ₄ +Azotobacter
T ₁₄	T ₅ +Azotobacter
T ₁₅	T ₃ +PSB
T ₁₆	T ₄ +PSB
T ₁₇	T ₅ +PSB
T ₁₈	T ₃ +Azotobacter+PSB
T ₁₉	T ₄ +Azotobacter+PSB
T ₂₀	T ₅ +Azotobacter+PSB

Sample Collection

Fresh leaf sample from each treatment were collected and bought to the laboratory for analysis of different parameters.

Biochemical Parameters

Estimation of Chlorophyll content

Chlorophyll content of the leaves was determined by procedure described by Hiscox and Israelstam, 1979. The chlorophyll a and b content of leaf was computed using formula suggested by Arnon (1949).

$$\text{Chlorophyll 'a' (mg/g)} = \frac{12.7(A663) - 2.69(A645)}{1000} * V$$

$$\text{Chlorophyll 'b' (mg/g)} = \frac{22.9(A645) - 4.68(A663)}{1000} * V$$

$$\text{Total Chlorophyll (mg/g)} = \frac{20.2(A645) + 8.02(A663)}{1000} * V$$

Physiological Parameters

Relative water content

The water potential of the leaf tissue was estimated according to the method of Slatyer (1955) [33]. About 1 gm of fresh leaves sample was weighed and dipped in 20 ml distilled water for three hour. After three hours turgid weight of the leaves was taken and then kept for drying in 60°C in Hot air oven till the weight become constant. The RWC was computed as

$$\text{RWC\%} = \frac{\text{Fresh weight of leaf} - \text{Dry weight of leaf}}{\text{Turgid weight} - \text{Dry weight}} * 100$$

Electrical Conductivity of leaf

The electrical conductivity of the leaf was estimated to the method of Bower and Wilcox, 1965. About 1 gm of fresh leaf was taken than dipped in 50ml distilled water for 10min. After 10 min EC was measured using electrical conductivity meter and then kept in water bath at 60°C for 10 min than again EC was measured.

Statistical Analysis

The data were analysed using Descriptive Statistics and SPSS20. Significant difference among mean were distinguished according to the Duncan's Multiple Test Range (Duncan, 1955) [10].

Result and Discussions

Physiological Character

a. Relative Water Content

The relative water content was seen to be significantly affected by different treatment. The Relative Water Content ranged from 72.646-87.73% with highest(85.2%) in T₁₇ was applied with 100% RDF through fertilizer+ 2kg Vermicompost+ Phosphate Solubilizing Bacteria(PSB) followed by T₁₈(85.02%) and T₃(84.99%). It was found that the RWC affect the chlorophyll pigment. Leaf pigment get damaged by water deficit as reported by (Montag U and Woo, 1990; Nilsen and orcutt, 1996) [25], due to production of Reactive Oxygen Species (ROS) such as O₂⁻ and H₂O₂ lead to lipid peroxidation and thus chlorophyll destruction (Mirnoff, 1993; Foyer *et al.*, 1994) [21, 12]. The chlorophyll is destroyed by water deficit and formation also prevented (Lessani and Mojtahedi, 2002) [18].

Table 2: Showing the effect of different treatment on RWC

Treatments	Relative water content (%)	
	Mean	Rank
T1	75.61 ^h	20
T2	76.95 ^h	19
T3	84.99 ^c	3
T4	77.77 ^h	18
T5	84.08 ^c	5
T6	80.48 ^{ef}	11
T7	78.33 ^g	15
T8	79.75 ^f	12
T9	83.83 ^d	7
T10	78.22 ^{gh}	17
T11	81.85 ^d	8
T12	81.16 ^{de}	10
T13	79.7 ^f	14
T14	83.88 ^c	6
T15	79.7 ^f	13
T16	81.48 ^d	9
T17	85.2 ^a	1
T18	85.02 ^b	2
T19	78.32 ^g	16
T20	84.31 ^c	4
Mean	81.0315	
S.Em±	0.4306	
Range	72.646-87.731	
CD (95.0%)	0.8617	

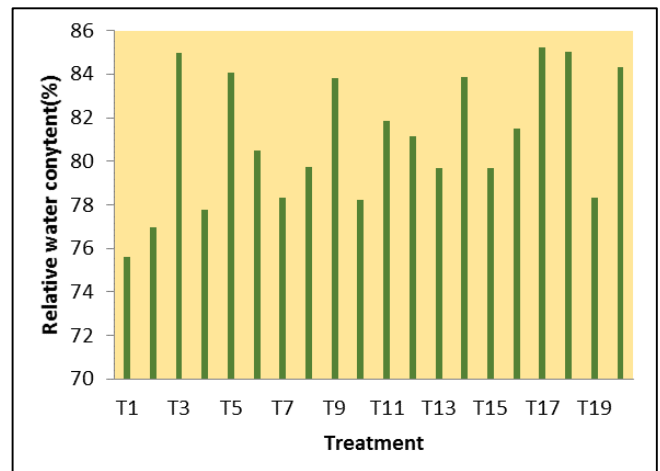


Fig 1: Influence of different treatment on RWC

b. Electrical Conductivity of leaf

The EC content of the leaf also found to be significantly affected by different nutrient treatment application. The EC content of leaf ranged from 0.03097-0.0979dSm⁻¹. The EC content of T₁₉ was recorded highest (0.09405dSm⁻¹) followed by T₁₈ (0.0922dSm⁻¹) and T₅ (0.0828dSm⁻¹). After 10 min of water bath treatment the EC of the leaf ranged from 0.05456-0.2527dSm⁻¹ with highest recorded in T₁₉ (0.245 dSm⁻¹) followed by T₆ (0.230 dSm⁻¹) and T₁₈ (0.2115 dSm⁻¹). The leaf chlorophyll content are also get affected by EC content. The chlorophyll content increase gradually as the EC content increase and decrease with lower EC content (Ding X *et al.*, 2018) [11].

Table 3: Effect of different treatment on EC content

Treatments	EC	EC after 10min
T1	0.0325	0.05628
T2	0.03265	0.0557
T3	0.0629	0.1774
T4	0.037	0.0673
T5	0.0828	0.1721
T6	0.077	0.230
T7	0.03247	0.08525
T8	0.07325	0.1932
T9	0.034	0.06335
T10	0.0374	0.0866
T11	0.04905	0.0672
T12	0.03795	0.07855
T13	0.03635	0.0603
T14	0.042	0.0805
T15	0.0429	0.0837
T16	0.03905	0.07315
T17	0.0391	0.0989
T18	0.0922	0.2115
T19	0.09405	0.245
T20	0.0807	0.19155
Mean	0.0527	0.1188
S.Em±	0.0027	0.00837
Range	0.03097-0.0979	0.05456-0.2527
CD (95.0%)	0.00559	0.01675

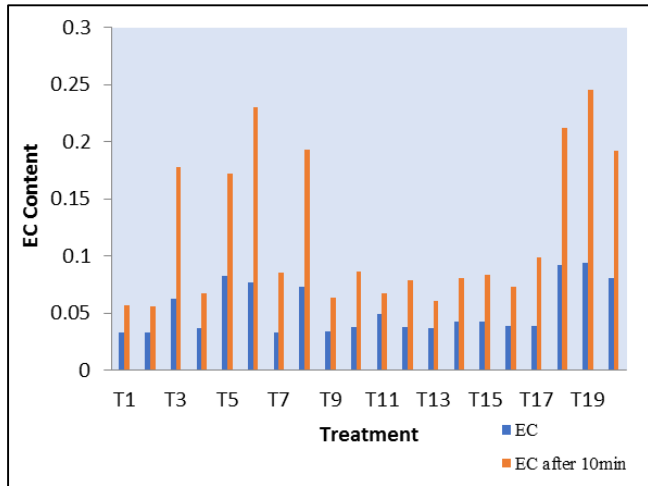


Fig 3: Influence of different treatment on EC content

Biochemical Characters

a. Chlorophyll content

Effect of inorganic, organic and biofertilizer were significant

on leaf chlorophyll content. Relative water content and Electrical Conductivity were found to effect significantly the Chlorophyll content. Chlorophyll 'a' content was higher in T₁₆ (4.70 mg/g) which was applied with 75%RDF through fertilizer+25% through vermicompost+ Phosphate Solubilizing Bacteria(PSB) over control T₁(1.05 mg/g). Chlorophyll 'b' content recorded high in T₇(3.18 mg/g) was treated with 75% RDF + *Azotobacter*. Total chlorophyll content of fresh leaf ranged from 1.719-8.049 mg/g with highest value in T₇ (7.68mg/g) followed by T₁₆ (7.55mg/g). The increase in chlorophyll content due to fertilization was also reported by (Amany S. Al-Erwy *et al.*, 2016). Fertilization with different manure increases the N content in the soil. Which helps in good N supply to the plant. Nitrogen is a important structural element of chlorophyll and protein molecules thus affecting the formation of chloroplasts and chlorophyll accumulation (Ray Tucker, 2004) [27]. Stability of chlorophyll in plants is affected by nitrogen concentration and is the important mineral in chlorophyll biosynthesis. Excesses of nitrogen decreases chlorophyll content in leaf (Bojovic and Stojanovic, 2005) [8].

Table 4: Chlorophyll content after different treatment

Treatments	Chl 'a' (mg/g)	Chl 'b' (mg/g)	Total Chl (mg/g)
T ₁	1.05	0.66	1.72
T ₂	2.76	2.38	5.14
T ₃	2.69	2.27	4.97
T ₄	1.11	0.77	1.99
T ₅	2.16	1.92	4.07
T ₆	2.03	1.73	3.76
T ₇	4.49	3.18	7.68
T ₈	2.39	2.12	4.51
T ₉	3.09	2.06	5.15
T ₁₀	2.65	2.57	5.31
T ₁₁	2.29	2.20	4.49
T ₁₂	3.46	2.51	5.97
T ₁₃	2.96	2.52	5.48
T ₁₄	2.78	2.06	4.85
T ₁₅	2.13	1.73	3.86
T ₁₆	4.70	2.84	7.55
T ₁₇	4.24	2.66	6.91
T ₁₈	3.74	2.33	6.06
T ₁₉	3.88	2.64	6.52
T ₂₀	4.66	2.64	7.29
Mean	2.96	2.189	5.164
S.Em±	0.136	0.079	0.2081
Range	1.041-4.761	0.6368-3.313	1.719-8.049
CD (95.0%)	0.273	0.1584	0.4164

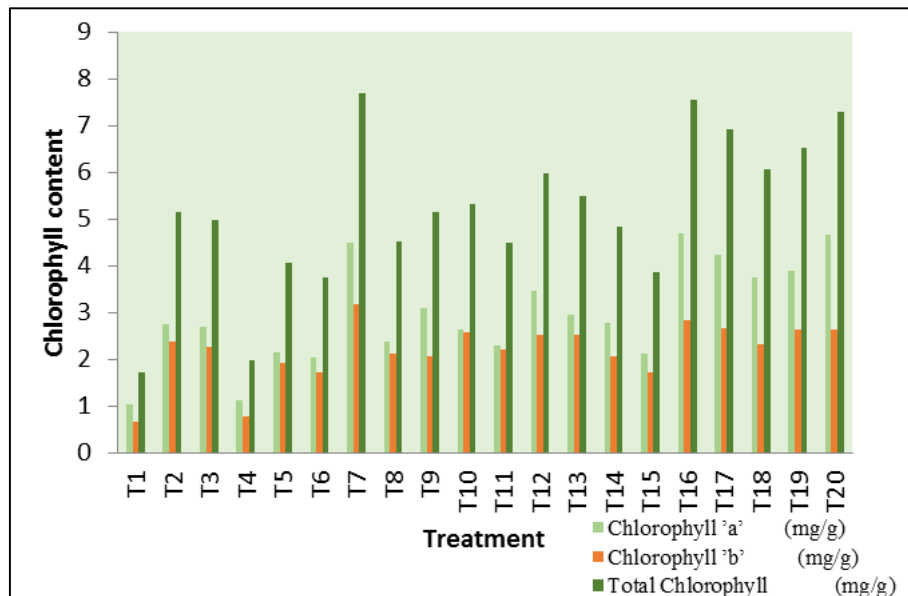


Fig 4: Influence of different treatment on chlorophyll content

Conclusion

From the above data collected it can be concluded that application of inorganic, organic and biofertilizer significantly influenced the leaf chlorophyll content, Relative water and EC content than the untreated plant. Combined application of all three fertilizer are most effective in increasing the nutrient content of leaf. Therefore, combined application of fertilizer may be better than the use of chemical fertilizer alone. Which shows negative effect on soil health and environment.

Acknowledgment

The authors like to thank the director of Central Tasar Research & Training Institute for providing the field facility and also former and present head of Soil Science and Chemistry Dr. P.P. Srivastava and Dr. D.M. Tiwary for providing the lab facility and also the head of University Department of Botany for courage and guidance.

References

- Ahmed MA, Ahmed AG, Mohamed MH, Rawfih MM. Integrated effect of organic Biofertilizers on wheat productivity, in new reclaimed sandy soil. Research Journal of Agriculture and Biological Sciences. 2011; 7(1):105-114.
- Allison FE. Soil Organic matter and its role in crop production. Amserdam K Scientific Publishing co., 1973; 16-23.
- Amany S Al-Erwy, Abdul moneam Al-Goukhy, Sameera O Bafeel. Effect of Chemical, Organic and Bio-fertilizers on Photosynthetic pigments, Carbohydrates and Minerals of Wheat (*Triticum aestivum* L.) Irrigated with Sea water. Int. J Adv. Res. Biol. Sci. 2016; 3(2):296-310.
- Amujoyegbe BJ, Opabode JT, Olayinka A. Effect of organic and inorganic fertilizer on yield and chlorophyll content of Maize (*Zea mays* L.) and Sorghum (*Sorghum bicolor* (L.) Moench). African Journal of Biotechnology. 2007; 6(16): 1869-1873.
- Amon OI. Copper enzymes in isolated chloroplasts polyphenoxidase Beta Vulgaris.Pl. Physiol. 1949; 24:1-15.
- Bhatia KN, Tyagi MP. Trueman's Elementary Biology. Trueman Book Company. Adda Hoshiarpur, Jalandhar. 2006; 2:U1-83-U1-85.
- Bhatia KN, Parashar AN. Plant Physiology. Trueman Book Company. Jalandhar City. 1997, 245-281.
- Bojovic, B. and Stojanovic, J. Chlorophyll and carotenoid content in wheat cultivars as a function of mineral nutrition. Arch. Biol. Sci., Belgrade, 2005; 57(4):283-290.
- Bower CA, Wilcox LV. Soluble Salts. In: Black, C.A., et al., Methods of Soil Analysis, American Society of Agronomy, Madison, 1965, 933-940.
- David B. Duncan. Multiple Range and Multiple T Tests. International Biometric Society. 1955; 11(1):1-42. <https://doi.org/10.2307/3001478>.
- Ding X, Jiang Y, Zhao H, Guo D, He L, Liu F, et al. Electrical conductivity of nutrient solution influenced photosynthesis, quality and antioxidant enzyme activity of Pak choi (*Brassica campestris* L. ssp. Chinensis) in a hydroponic system. PLoS ONE. 2018; 3(8):e0202090. <https://doi.org/10.1371/journal.pone.0202090>.
- Foyer CHO, Decourvieres, KJ Kunert. Photo-oxidative stress in Plants. Plant Physiol. 1994; 92:696-717.
- Functioning of central silk board & performance of Indian Silk Industry Central Silk Board (Ministry of Textiles, Govt. of India). 2021, Bangalore-560068.
- Hiscox JD, Israelstam GF. A method for the estimation of chlorophyll from leaf tissue without maceration. Can J Bot. 1979; 57:1332-1334.
- Kaiser WM. Effects of water deficit on photosynthetic capacity. Physiologic plantarum. 1987; 71:142-149.
- Kozai T, Niu G, Takayahi M. Plant factory as a resource efficient closed plant production system: In An indoor vertical farming system for efficient quality food production. Academic press Cambridge, MA, USA, 2015, 69-90.
- Le Bot J, Adamowicz S, Robin P. Modelling plant nutrition of horticulture crops: A review. Sci. Hortic. 1998; 74:47-82.
- Lessani H, M Mojtahedi. Introduction to plant Physiology (Translation).6th edn, Tehran University Press, Iran, ISBN: 964-03-3568-1, 2002; pp: 726.

19. Lu N, Bernardo EL, Tippayadarapanich C, Takagahi M, Kagawa N, Yamori W. Growth and accumulation of secondary metabolites in *Perilla* as affected by photosynthetic photon flux density and electrical conductivity of the nutrient solution. *Font. Plant Sci.* 2017; 8:708.
20. M Hassanzadeh, A Ebadi, M Panahyan-e-kivi, AG Eshghi, Sh Jamaali-e-Somarin. Evaluation of Drought stress on Relative Water Content and Chlorophyll content of Sesame (*Sesamum indicum* L.) Genotypes at early flowering stage. *Research Journal of Environmental Sciences.* 2009; 3(3):345-350.
21. Mirnoff N. The role of active oxygen in the response of Plants to water deficit and desiccation. *New Phytol.* 1993; 125:27-58
22. Montagu KD, KC Woo. Recovery of tree photosynthetic capacity from seasonal drought in the wet dry tropics: The role of phyllode and canopy processes in *Acacia auriculiformis*. *Aust. J Plant Physiol.* 1999; 26:135-145.
23. Nageswara RRC, HS Talwar GC. Wright Rapid assessment of specific leaf area and leaf nitrogen in peanut (*Arachis hypogaea* L.) using chlorophyll meter. *J Agron. Crop Sci.* 2001; 189:175-182.
24. Nemail KS, Van Iersel MW. Light intensity and fertilizers concentration: I. Estimating Optimal Fertilizer Concentrations from Water-use Efficiency of Wax Begonia. *Hort science.* 2004; 39(6):1287-1292.
25. Nilsen ET, DM Orcutt. *Physiology of plant under stress abiotic factors* 2nd edn., John Wiley and Sons. Inc., New York. ISBN:0471170089. 1996; pp: 689.
26. Purwanto Budi Santosa, Junaidah junaidah Effect of the combination of some fertilizers on chlorophyll content of Gemor (*Nothaphoebe coriacea*), Indonesia. *Asian Journal of Applied Sciences.* 2015; 3(4):699-703.
27. Ray tucker M. *Primary Nutrients and Plant Growth*, In: *Essential Plant Nutrients*, North-Carolina, Department of Agriculture, 2004.
28. Roy DK, Singh BP. Effect of level and time of Nitrogen application with and without vermicompost on yield, yield attributes and quality of malt barley (*Hordeum vulgare*). *Indian Journal of Agronomy.* 2006; 51:40-42.
29. Sabashini HD, Malavannan S, Kumar P. Effect of biofertilizers on yield of rice cultivars in Pondicherry, India. *Asian Journal of Agriculture Research.* 2007; 1(3):146-150.
30. Schonfeld MA, RC Johnson, BT Carwer, DW Mornhinweg. Water relations in winter wheat as drought resistance indicators. *Crop Sci.* 1988; 28:526-531.
31. Siavoshi M, Laware SL. Role of organic fertilizers on chlorophyll content in rice (*Oryza sativa* L.). *Trends in Life Sciences.* 2013; 2(3):13-17.
32. Signore A, Serio F, Santamaria P. A targeted management of the nutrient solution in a soilless tomato crop according to plant needs. *Front Plant Sci.* 2016; 7:391. <https://doi.org/10.3389/fpls.2016.00391> PMID:27242804.
33. Slatyer RD. Studies of the water relations of crop plant grown under natural rainfall in Northern Australia. *Aust. J Agr. Res.* 1955; 6:365-377.
34. Sonneveld C, Voogt W. *Plant nutrition of greenhouse crops*, Springer, ISBN9048125316, New York, U.S.A. 2009.
35. United States Department of Agric. Improving soil with organic waste report to congress in respond to section 1416 of Food and Agriculture Act of 1977 (pl-95-113) US Government Printing Office, Washington DC. 1978.
36. Vijay Paul, Laxmi Sharma, Rajeev Kumar, Rakesh Pandey and R.C. Meena. Estimation of chlorophylls/photosynthetic pigments. Their stability is an indicator of crop plant tolerance to abiotic stresses. *Manual of ICAR Sponsored Training Programme on "Physiological Techniques to analyze the Impact of Climate Change on crop plants.* Division of Plant Physiology, IARI. New Delhi, 2017.
37. Wali Asal MA. The combined effect of mineral organic and bio-fertilizers on the productivity and quality of some wheat cultivars. Ph.D. Thesis. Faculty of Agricultural Alex University, Egypt, 2010.
38. Wright GC, RC Nageswara Rao, GD Farquhar. Water use efficiency and carbon isotope discrimination in peanut under water deficit conditions. *Crop Sci.* 1994; 34:92-97.
39. www.imotforum.com/2017/11/silk-farming-agripreneurs.